

## Effects of indomethacin on renal hemodynamics, urinary enzymes and thromboxane B<sub>2</sub> following envenomation of Russell's viper in dogs.

Sopit Thamaree\*

Narongsak Chaiyabutr\*\* Smai Leepipatpaiboon\*\*\*

Kanit Buranasiri† Piyaral Tosukowong†

Piroj Sirivongs\* Visit Sitprijia††

Thamaree S, Chaiyabutr N, Leepipatpaiboon S, Buranasiri K, Tosukowong P, Sirivongs P, Sitprijia V. Effects of indomethacin on renal hemodynamics, urinary enzymes and thromboxane B<sub>2</sub> following envenomation of Russell's viper in dogs. *Chula Med J* 1987 May; 31(5) : 387-398

*The effects of indomethacin (5 mg/kg, i.v.) on renal hemodynamics, urinary enzymes, plasma and urinary thromboxane B<sub>2</sub> (TxB<sub>2</sub>) following a 40 min intrarenal arterial infusion of Russell's viper venom (RVV, 1.25 ug/kg/min) were studied in anesthetized male mongrel dogs. All parameters were determined before and at every 20 min after envenomation. Urinary enzymes were assayed by colorimetric method. Plasma and urinary thromboxane B<sub>2</sub> (P<sub>TxB<sub>2</sub></sub>, U<sub>TxB<sub>2</sub></sub>) were measured by radioimmunoassay after extraction and purification steps. The reduction of mean arterial pressure (MAP) and the increase of urine flow rate (V), P<sub>TxB<sub>2</sub></sub> and U<sub>TxB<sub>2</sub></sub> induced by RVV were inhibited by indomethacin pretreatment. Renal blood flow (RBF) and glomerular filtration rate (GFR), relatively unchanged by RVV, were markedly reduced by indomethacin pretreatment. Only urinary N-acetyl-β-D-glucosaminidase (NAG) was increased after envenomation. These results suggest an augmented synthesis of TxB<sub>2</sub> and/or disturbance of active tubular reabsorption of U<sub>TxB<sub>2</sub></sub>. After the inhibition of prostaglandin synthesis by indomethacin, the reduction of renal function and hypertensive response to RVV may be accounted for by the renin-angiotensin system. The elevation of U<sub>NAG</sub> indicates a direct nephrotoxicity of RVV.*

Reprint requests : Thamaree S. Department of Pharmacology, Faculty of Medicine, Chulalongkorn University, Bangkok 10500, Thailand.

Received publications. February 9, 1987.

\* Department of Pharmacology, Faculty of Medicine, Chulalongkorn University.

\*\* Department of Physiology, Faculty of Veterinary Science, Chulalongkorn University.

\*\*\* Department of Obstetrics and Gynecology, Faculty of Medicine, Chulalongkorn University.

† Department of Biochemistry, Faculty of Medicine, Chulalongkorn University.

†† Department of Medicine, Faculty of Medicine, Chulalongkorn University.

โสภิต ธรรมอารี, ณรงค์ศักดิ์ ชัยบุตร, สมัย ลีพิพัฒน์ไพบูลย์, ชนิษฐ บวรณศิริ, ปิยะวัฒน์ โดสุโขวงศ์, ไพโรจน์ ศิริวงษ์, วิศิษฎ์ สิตปรีชา. ฤทธิ์ของอินโดเมธธาซินต่อการทำงานของไต, เอ็นซิมในปัสสาวะ และรอมบ็อกเซน บีทู ภายหลังจากการได้รับพิษงูแมวเซาในสุนัข. จุฬาลงกรณ์เวชสาร 2530 พฤษภาคม; 31(5) : 387-398

คณะผู้วิจัยได้ศึกษาฤทธิ์ของพิษงูแมวเซาที่ฉีดเข้าหลอดเลือดแดงของไตเป็นเวลา 40 นาที ในขนาด 1.25 ไมโครกรัม/กิโลกรัม/นาที ที่มีผลต่อการทำงานของไต, เอ็นซิมในปัสสาวะ และระดับรอมบ็อกเซน บีทู ในพลาสมา และปัสสาวะของสุนัขที่ได้รับการฉีดอินโดเมธธาซินเข้าหลอดเลือดดำที่ขาในขนาด 5 มิลลิกรัม/กิโลกรัม (7 ตัว) ก่อนการฉีดพิษงู 40 นาที เปรียบเทียบกับสุนัขที่ไม่ได้รับอินโดเมธธาซินมาก่อน (7 ตัว) โดยวัดผล ทุก ๆ 20 นาที ภายหลังจากเริ่มให้พิษงูแมวเซา แอ็คติวิตีของเอ็นซิมในปัสสาวะวัดโดยวิธีคลอริเมตริก ระดับรอมบ็อกเซน บีทู ในพลาสมา และปัสสาวะวัดโดยวิธีเรดิโออิมมูโนแอสเซย์ภายหลังขั้นตอนการสะกัดและทำให้บริสุทธิ์แล้ว พิษงูแมวเซาทำให้ความดันเลือดลดลง เพิ่มการขับปัสสาวะและเพิ่มระดับรอมบ็อกเซน บีทู ในพลาสมา และปัสสาวะ ผลเหล่านี้กลับเป็นตรงกันข้ามเมื่อสุนัขได้รับอินโดเมธธาซินก่อนได้รับพิษงู เลือดที่ไปเลี้ยงไตและอัตราการกรองที่โกลเมอรูลัส ไม่ค่อยเปลี่ยนแปลงเมื่อได้รับพิษงูแต่จะลดลงเมื่อสุนัขได้รับอินโดเมธธาซินมาก่อน เอ็นซิมในปัสสาวะที่มีการเพิ่มขึ้นอย่างเด่นชัดภายหลังได้รับพิษงูมีเพียงเอ็น-อะเซทิล-บีต้า-ดี-กลูโคซามินิเดส (เอ็นเอจี) ชนิดเดียว ผลเหล่านี้แสดงให้เห็นว่าพิษงูแมวเซาทำให้มีการสังเคราะห์รอมบ็อกเซน บีทู เพิ่มขึ้น และ/หรือมีการรบกวนการดูดซึมกลับของ รอมบ็อกเซน บีทู ที่ท่อไต การที่พิษงูทำให้ความดันเลือดสูงขึ้นภายหลังการยับยั้งการสังเคราะห์ พรอสตาแกลนดินส์แล้วน่าจะเป็นผลของระบบเรนิน-แองจิโอเทนซินที่เด่นชัดขึ้นมา เอ็นซิมเอ็นเอจีเพิ่มขึ้น อาจเป็นสิ่งที่ชี้แนะว่าพิษงูแมวเซามีพิษโดยตรงต่อไตก็ได้.

Russell's viper venom (RVV) is known to cause renal damage including tubular necrosis<sup>(1,2)</sup>, cortical necrosis<sup>(3)</sup> and glomerulonephritis.<sup>(4)</sup> Acute renal failure (ARF) which is recognized to be an important cause of death in patients bitten by Russell's viper may partly result from tubulotoxicity of this venom. It has been suggested that a wide variety of enzymes normally excreted into the urine are increased in various disease states including in renal tubular cell injury.<sup>(5)</sup> The early work of Wellwood et al.<sup>(6)</sup> had indicated that certain lysosomal enzymes from the proximal tubular cells : N-acetyl-beta-D-glucosaminidase (NAG) and beta-galactosidase (GAL), or from the brush border : gamma-glutamyl-transpeptidase ( $\gamma$ GT) and alanine-amino-peptidase (AAP) are early markers of tubular damage induced by aminoglycosides. There was no data on enzymuria despite clinical evidence suggesting a direct tubulotoxicity of RVV.

Furthermore, the reduction in renal hemodynamics clearly observed in experimental dogs envenomated with RVV<sup>(7)</sup>, may also play a role in the initial pathogenesis, in the ischemic model of ARF<sup>(8)</sup>. It has been proposed that catecholamine release, renin-angiotensin activation<sup>(7)</sup> and prostaglandin release<sup>(9)</sup> may be responsible for the impairment of renal function.

In this study the effects of Russell's viper venom on renal hemodynamics, urinary enzymes, plasma thromboxane B<sub>2</sub> (P<sub>TxB<sub>2</sub></sub>) and urinary thromboxane B<sub>2</sub> (U<sub>TxB<sub>2</sub></sub>) in dogs pretreated with indomethacin compared to non-pretreated dogs were investigated in an attempt to elucidate the relationship between the nephrotoxicity of RVV and urinary enzymic activities, the role of thromboxane B<sub>2</sub> in renal hemodynamic changes and effects of indomethacin, a cyclooxygenase inhibitor.

### Material and Method

Fourteen mongrel dogs weighing 10 to 15 kg were anesthetized by intravenous injection of thiopentone sodium at the dosage of 20-25 mg/kg. Supplementary doses were given as necessary to maintain light anesthesia. A femoral artery was cannulated for arterial blood sample collection and for recording of arterial blood pressure and heart rate. The heart rate was recorded on a Grass physiograph. Renal artery was cannulated for venom infusion. Femoral vein was cannulated for fluid infusion. A polyethylene catheter was introduced into the bladder for urine collection.

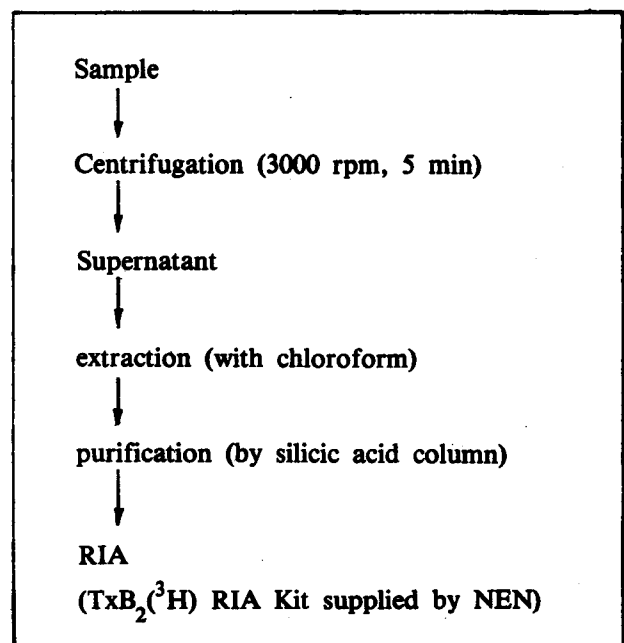
Renal plasma flow and glomerular filtration rate were measured by p-aminohippurate and inulin clearances using standard techniques.<sup>(10)</sup> Renal

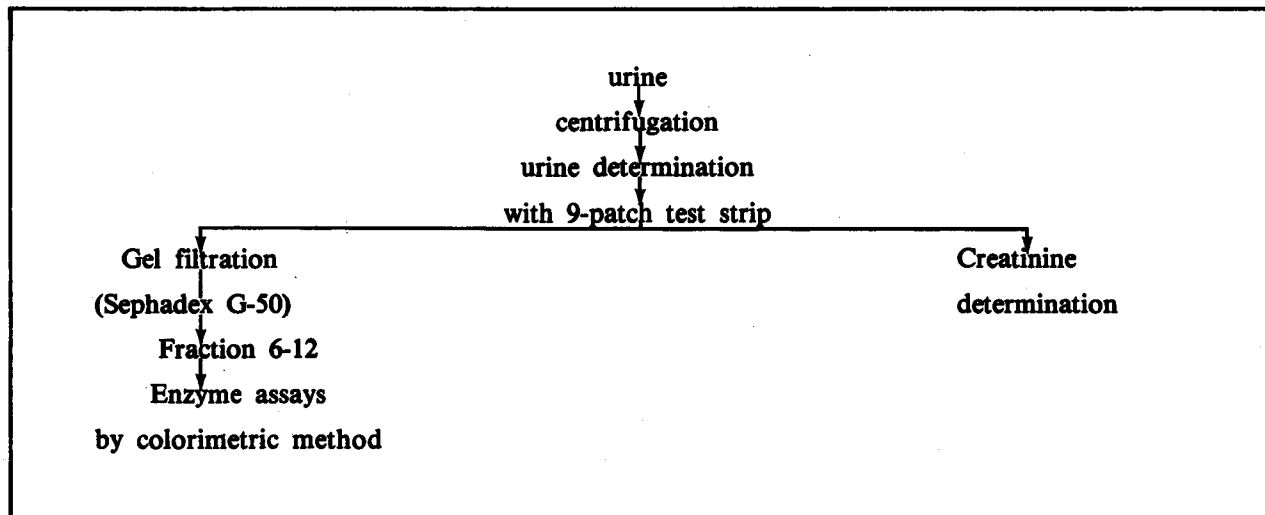
blood flow was calculated from p-aminohippurate clearance and packed red cell volume. Plasma levels of p-aminohippurate and inulin were maintained approximately at 0.03 mg/ml and 0.20 mg/ml by sustained infusion of isotonic saline solution containing p-aminohippurate (Merck, Sharpe and Dohme) and inulin (Sigma Chemical Co.) following the priming doses. After 40 min of equilibration, urine was collected for 20 min and an arterial blood sample was obtained at the midpoint of urine collection. This served as the control period. Thence, Russell's viper (*Vipera russelli siamensis*) venom, donated by the Science Division of the Thai Red Cross Society, was given by intrarenal arterial infusion at a dose of 0.05 mg/kg. The lyophilized venom was dissolved in 20 ml of 0.9% sodium chloride solution (NSS). The infusion rate was 0.5 ml/min.

For the indomethacin pretreated group, indomethacin was administered intravenously at a dose of 5 mg/kg, 40 min before envenomation of RVV. Blood sampling and urine collection were made twice over a period of 40 min, after indomethacin pretreatment and at every 20 min after envenomation.

The analysis of urine and blood samples were performed of p-aminohippurate and inulin,<sup>(10)</sup> sodium and potassium (flame photometric method), chloride (by chloridometer), TxB<sub>2</sub> (as shown in diagram I). Urinary lysosomal enzyme activities were measured by colorimetric method<sup>(11-13)</sup> as shown stepwise in diagram II.

Diagram I Steps of TxB<sub>2</sub> assay.



**Diagram II** Steps of analysis of urinary enzyme activities.**Result**

The reduction in arterial blood pressure after envenomation was inhibited by indomethacin pretreatment. Heart rate was slightly decreased in both groups after envenomation (Table 1.) Packed red cell volume was not altered by indomethacin pretreatment but was slightly increased in response to RVV. Renal blood flow (RBF) and renal plasma flow (RPF) were slightly increased during the first 20 min after envenomation but gradually decreased toward the control level, while indomethacin caused a significant decrease in RBF and RPF with further decreases after envenomation. Glomerular filtration rate (GFR), relatively unchanged by RVV alone, was decreased slightly after indomethacin pretreatment and significantly following envenomation. The rising of urine flow rate (V) by 15-65% in response to RVV was inhibited by indomethacin pretreatment (Fig. 1). Urinary excretion of sodium ( $U_{Na} V$ ), chloride ( $U_{Cl} V$ ) and potassium ( $U_K V$ ), and the fractional excretion of sodium ( $FE_{Na}$ ) and chloride

( $FE_{Cl}$ ) were increased during the first 20 min of envenomation parallel to the increase in urine flow rate and then declined toward the control level. Indomethacin pretreatment tended to decrease the urinary excretion of sodium, chloride and potassium which gradually increased toward the control level during a 40-min of envenomation. Fractional excretion of sodium, chloride and potassium rose throughout the experimental period but not significantly different from the control (Table 2). The urinary enzyme activity of NAG, significantly increased after the end of a 40-min intrarenal arterial infusion of RVV, was elevated slightly by indomethacin pretreatment and again significantly during envenomation. Other urinary enzyme activities fluctuated but showed a tendency toward an increase after envenomation (Fig. 2). A 7 to 10 fold elevation of urinary  $TxB_2$  was observed during envenomation. Plasma  $TxB_2$  followed a parallel elevation. Indomethacin completely inhibited the rise in  $P_{TxB_2}$  and  $U_{TxB_2}$ . (Fig. 3, 4)

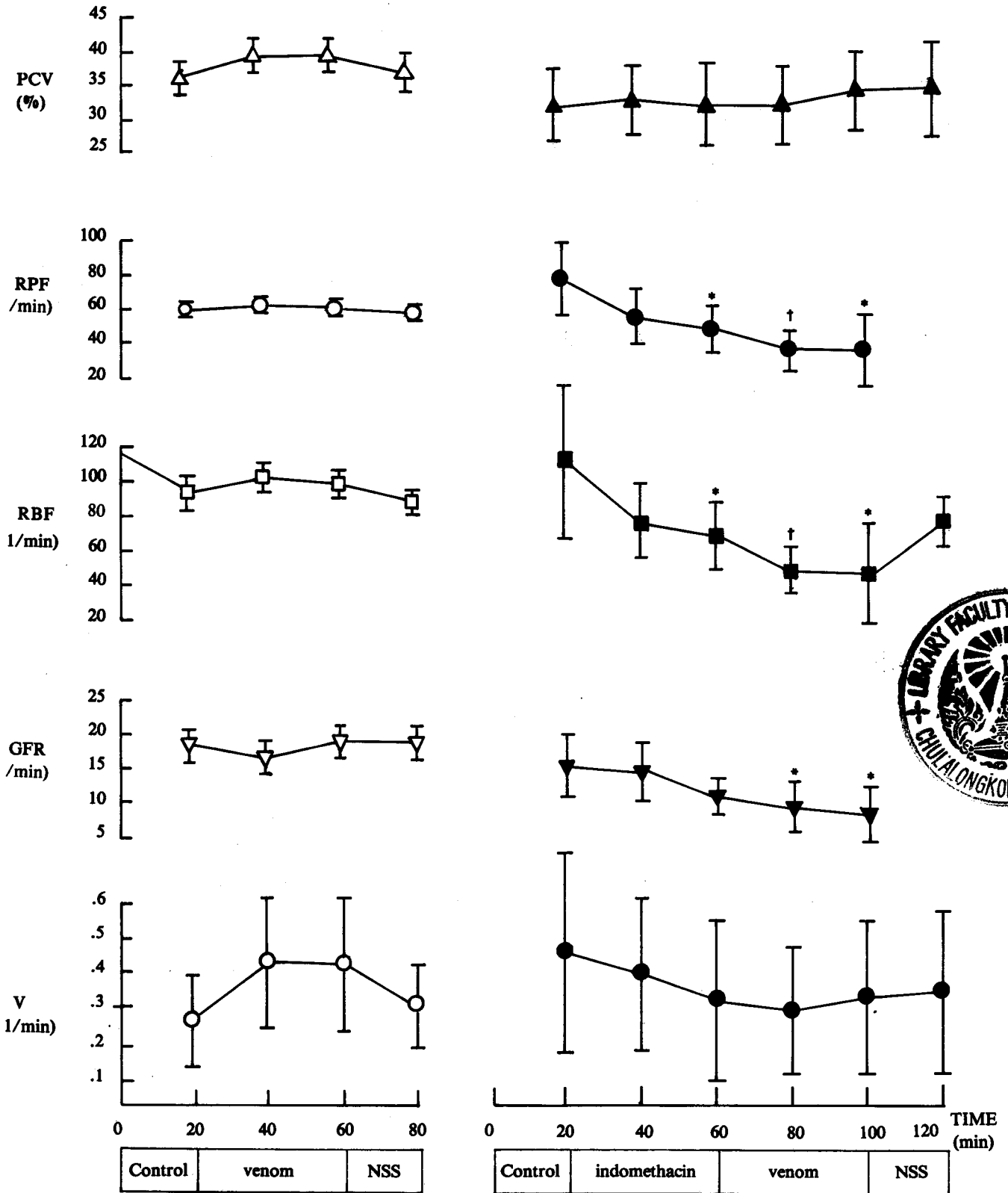
**Table 1** Effect of Russell's viper venom on blood pressure and heart rate.

Parameter	Control	Indomethacin		Russell's viper venom		NSS 20 min
		20 min	40 min	20 min	40 min	
Mean arterial pressure (mm Hg)	128 ± 8	not given	not given	118 ± 10**	118 ± 12	117 ± 11
	122 ± 24	128 ± 22*	130 ± 19*	129 ± 22*	137 ± 17*	140 ± 17†
Heart rate (beats/min)	154 ± 11	not given	not given	147 ± 9	135 ± 10	135 ± 12
	145 ± 21	130 ± 17	128 ± 15	127 ± 21	130 ± 25	134 ± 26

Values are mean ± S.D., n = 7

\*P < 0.05, \*\*P < 0.025, † P < 0.01 when compared to the control

NSS = Normal saline solution



**Figure 1** Effects of Russell's viper venom on packed cell volume (PCV), renal plasma flow (RPF), renal blood flow (RBF), glomerular filtration rate (GFR) and urine flow rate (V) in indomethacin non-pretreated (n=7) and pretreated dogs (n=7). Values are mean  $\pm$  S.D., \*P<0.05, †P<0.001 when compared to the control. NSS = normal saline solution.

Table 2 Effect of Russell's viper venom on sodium, chloride and potassium excretion

	Control	Indomethacin		Russell's viper venom		NSS 20 min
		20 min	40 min	20 min	40 min	
U <sub>Na</sub> V (mEq/min)	40.64 ± 40.52	not given	not given	53.34 ± 36.02	39.36 ± 25.26	29.75 ± 18.04
FE <sub>Na</sub> (%)	1.81 ± 2.17	not given	not given	3.25 ± 3.19*	1.73 ± 1.28	1.48 ± 1.01
U <sub>Na</sub> V (mEq/min)	48.4 ± 32.48	38.09 ± 41.14	35.85 ± 38.22	35.75 ± 25.88	41.50 ± 33.44	46.08 ± 37.52
FE <sub>Na</sub> (%)	2.2 ± 1.1	2.2 ± 1.7	3.3 ± 4.2	14.9 ± 30.1	4.2 ± 2.2	7.3 ± 3.8
U <sub>Cl</sub> V (mEq/min)	45.85 ± 45.62	not given	not given	59.36 ± 39.13	46.99 ± 30.69	29.84 ± 17.87
FE <sub>Cl</sub> V (%)	2.32 ± 2.86	not given	not given	4.33 ± 4.76*	2.52 ± 2.05	1.70 ± 1.18
U <sub>Cl</sub> V (mEq/min)	54.2 ± 50.12	51.19 ± 68.02	43.57 ± 65.71	46.31 ± 43.48	50.91 ± 51.73	54.04 ± 46.62
FE <sub>Cl</sub> V (%)	2.95 ± 2.25	3.4 ± 2.7	5.1 ± 8.0	5.2 ± 5.5	5.3 ± 3.1	9.2 ± 4.0
U <sub>K</sub> V (mEq/min)	11.25 ± 4.85	not given	not given	12.54 ± 4.54	9.78 ± 3.57	9.70 ± 2.09
FE <sub>K</sub> (%)	23.53 ± 15.05	not given	not given	32.16 ± 17.03*	20.15 ± 9.92	23.10 ± 8.59
U <sub>K</sub> V (mEq/min)	8.74 ± 7.07	6.25 ± 3.65	6.48 ± 4.07	6.65 ± 3.89	8.16 ± 5.21	8.11 ± 4.93
FE <sub>K</sub> (%)	18.45 ± 8.1	17.4 ± 9.2	25.3 ± 19.2	27.7 ± 18.0	47.6 ± 50.1	32.9 ± 13.1

Values are mean ± S.D., n = 7

UV = urinary excretion, FE = fractional excretion, Na = sodium, Cl = chloride, K = potassium.

\*P < 0.05 when compared to the control.

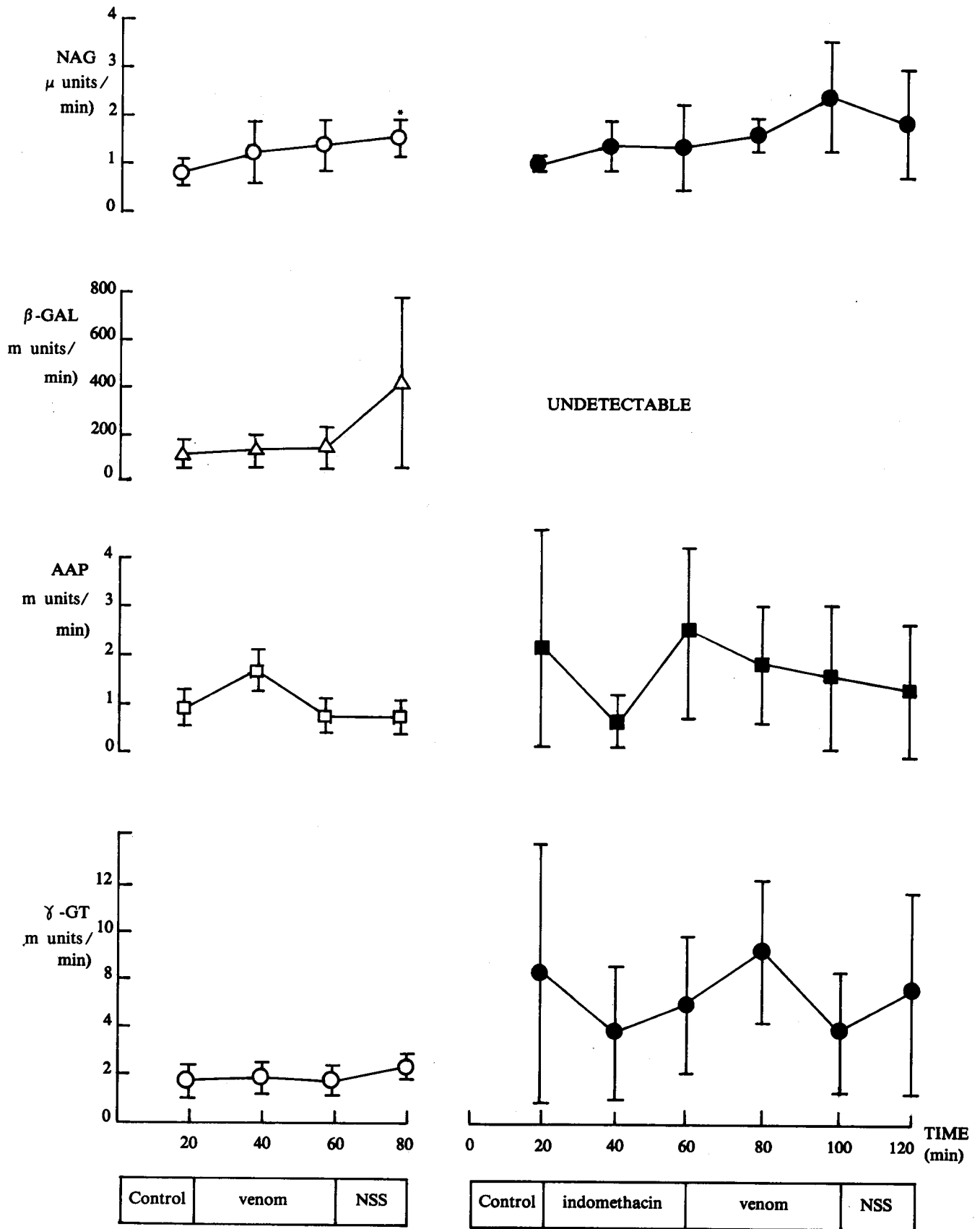
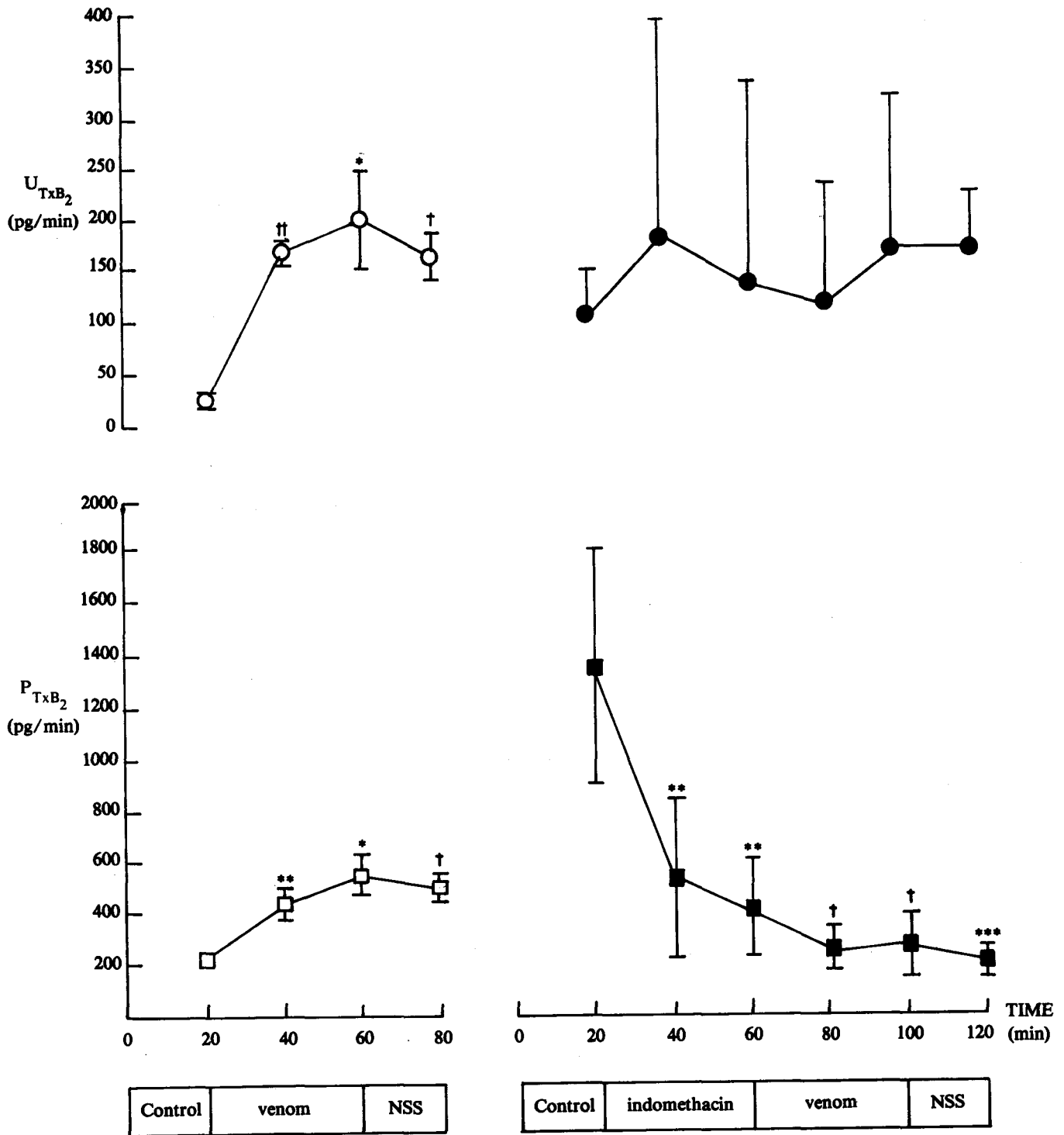
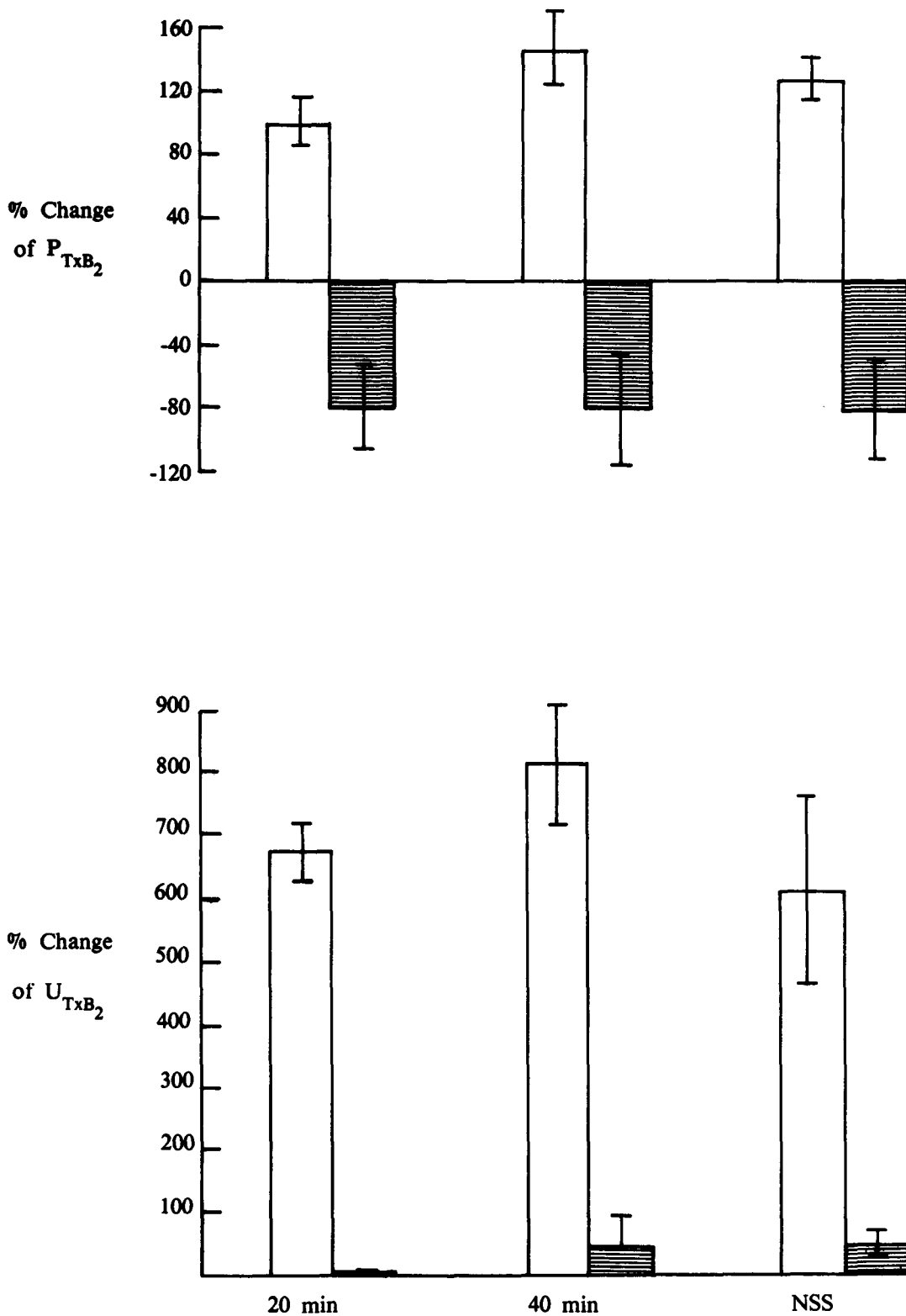


Figure 2 Effects of Russell's viper venom on urinary enzyme activities (n = 7). Values are mean  $\pm$  S.D., \*P < 0.05 when compared to the control. NSS = normal saline solution.



**Figure 3** Effects of Russell's viper venom on the urinary and plasma levels of thromboxane B<sub>2</sub> in the indomethacin non-pretreated (n = 7) and pretreated dogs (n = 7). Values are mean ± S.D., \*P < 0.05, \*\*P < 0.025, † P < 0.01, ‡‡‡P < 0.0005, †† P < 0.001 When compared to the control. NSS = normal saline solution.





**Figure 4** Comparing of the percentage changes from the control level of the plasma and urinary thromboxane B<sub>2</sub> (P<sub>TxB<sub>2</sub></sub> and U<sub>TxB<sub>2</sub></sub>) between the indomethacin non-pretreated □ and indomethacin pretreated ■ dogs after 20 min and 40 min of envenomation and a 20-min infusion of normal saline solution (NSS). Each bar is mean ± S.D. of 7 dogs.

## Discussion

The results from our studies show that Russell's viper venom directly infused via renal artery at the dose of 1.25  $\mu\text{g}/\text{kg}/\text{min}$  can slightly affect the systemic circulation and renal hemodynamics. The reduction of mean arterial blood pressure may be attributable to splanchnic vasodilatation.<sup>(14,15)</sup> It is believed that Russell's viper venom causes vasodilatation through its effect on prostaglandins and kinin release as well as other slow reacting substances.<sup>(9,16)</sup> Our results showed a parallel increase of  $P_{\text{TxB}_2}$  and  $U_{\text{TxB}_2}$  due to Russell's viper venom despite no marked change in hemodynamics. The increased  $P_{\text{TxB}_2}$  and  $U_{\text{TxB}_2}$  may have partly been the result of augmented synthesis of  $\text{TxB}_2$  in the renal vasculature. The vasoconstrictor action of  $\text{TxB}_2$  may be counteracted by other renal hormones including vasodilatory prostaglandins. Indomethacin, a cyclooxygenase inhibitor, given prior to envenomation can completely inhibit the rise in  $P_{\text{TxB}_2}$  which accompanied the marked increase in blood pressure in response to the venom. Activation of renin-angiotensin system may be responsible for the hypertensive effect of the venom since, in other study, angiotensin converting enzyme inhibitor has been shown to attenuate this effect<sup>(17)</sup>. The slight elevation of packed cell volume in both groups was presumably due to splenic contraction.<sup>(7)</sup> Although the renal blood flow and glomerular filtration rate were relatively unchanged after envenomation, it should be noted that urine flow rate was increased by approximately 15-65%, accompanied by an increase in urinary excretion of sodium, chloride, potassium and thromboxane  $\text{B}_2$  as well. They could be accounted for by a direct tubular dysfunction since thromboxane  $\text{B}_2$ , like uric acid, is actively reabsorbed from the proximal tubule.<sup>(8)</sup>

Recently it has been reported that Russell's viper venom induced depolarization of the proximal tubular cell possibly by 3 mechanisms.<sup>(19)</sup> It has been proposed that the failure of the proximal tubular reabsorption of sodium giving rise to a sodium overload in the distal tubular urine can induce the secretion of renin by the macula densa.<sup>(20)</sup> The activation of angiotensin could induce a decrease in the renal blood flow and cortical ischemia followed by a decrease in glomerular filtration rate. The reduction in renal blood flow may be modulated by the production of vasodilatory prostaglandins,  $\text{PGI}_2$  and  $\text{PGE}_2$ , in the kidney.<sup>(21)</sup> We did not detect any production of  $\text{PGI}_2$  and  $\text{PGE}_2$  but clearly observed a decreased renal blood

flow and glomerular filtration rate in response to Russell's viper venom after inhibition of prostaglandin synthesis by indomethacin. The activation of renin-angiotensin system could likely account for this. Our result may have reflected the role of angiotensin II on prostaglandin production which McGiff et al<sup>(22)</sup> were the first to demonstrate and was subsequently confirmed by many investigators.<sup>(21,23,24)</sup> Urine flow rate changes similar to renal blood flow and glomerular filtration rate during the first 20 min of envenomation then rose despite the decrease in renal blood flow and glomerular filtration rate. Urinary excretion of sodium, chloride and potassium seemed to have decreased relatively less than the decrease in glomerular filtration rate and therefore the fractional excretion of sodium, chloride and potassium were increased. This result suggested that the inhibition of tubular reabsorption of sodium presumably occurred at sites proximal to the distal nephron thus a definite amount of sodium reached this part of the nephron and a small amount under reabsorption in exchange to potassium. Aldosterone, responsible for sodium reabsorption in the distal nephron, may be increased primarily due to the increase in angiotensin II. The correlation between plasma aldosterone concentration and plasma renin activity has been described by Zadik and Kowarski,<sup>(25)</sup> that the decrease in plasma renin activity is most likely the primary cause of the lower plasma aldosterone concentration; possibly, the opposite may be true. Elevated level of NAG probably indicated the evidence of renal injury corresponding to other types of renal disease or injury previously reported.<sup>(26,27)</sup> The measurement of AAP,  $\gamma$ -GT or  $\beta$ -GAL appeared on the contrary, to have had little significance. In summary, Russell's viper venom at this dose may directly alter the active reabsorption in the tubular cells. Elevation of  $P_{\text{TxB}_2}$  and  $U_{\text{TxB}_2}$  indicated the augmentation of synthesis and/or alteration of tubular reabsorption. Activation of the renin-angiotensin system seemed to have been responsible for the decrease in the renal blood flow and glomerular filtration rate. Experimental data on plasma renin activity changes should be worked out. It remains to be determined whether the levels of prostacycline ( $\text{PGI}_2$ ),  $\text{PGE}_2$  and aldosterone are changed and if so, what role they may eventually play in renal hemodynamic changes. Renal histological changes should be performed in accompaniment with a study on urinary enzymes.

## Acknowledgement

This study was supported by the Rachadapiseksompoj-China Medical Board Grant. We gra-

tefully acknowledge the excellent technical assistance of Dr. C. Buranakarl

## References

1. Sitprija V, Benyajati C, Boonpucknavig V. Further observations on renal insufficiency in snakebite. *Nephron* 1974; 13 (5) : 396-403
2. Sitprija V, Boonpucknavig V. The kidney in tropical snakebite. *Clin. Nephrol.* 1977 Sep ; 8 (3) : 377-383
3. Chugh KS, Aikat BK, Sharma BK, Dash KC, Mathew MT, Das KC. Acute renal failure following snakebite. *Am J Trop Med Hyg* 1975 Jul; 24 (4) : 692-697
4. Sitprija V, Boonpucknavig V. Extracapillary proliferative glomerulonephritis in Russell's viper bite. *Br. Med. J.* 1980 Jun 14; 280 (6229) : 1417
5. Wilkinson JH. Diagnostic significance of enzyme determination in urine. In : Dubach UC, ed. *Current Problems in Clinical Biochemistry : Enzymes in Urine and Kidney.* Vol 2 Bern, Switzerland : Verlag Hans Huber, 1968. 207
6. Wellwood JM, Simpson PM, Tighe JR, Thompson AE. Evidence of gentamicin nephrotoxicity in patients with renal allografts. *Br Med J* 1975 Aug 2; 3 (5918) : 278-281
7. Tungthanathanich P, Chaiyabutr N, Sitprija V. Effect of Russell's viper (*Vipera russelli siamensis*) venom on renal hemodynamics in dogs. *Toxicon* 1986 Aug ; 24 (4) : 365-371
8. Schrier RW, Conger JD. Acute renal failure : pathogenesis, diagnosis, and management. In : Schrier RW, ed. *Renal and Electrolyte Disorders.* Boston : Brown, 1980. 375-408
9. Huang HC. Release of slow reacting substance from the guinea pig lung by phospholipase A<sub>2</sub> of *Vipera russelli* venom. *Toxicon* 1984 Mar ; 22 (3) : 359-372
10. Smith HW. *Principles of Renal Physiology.* New York : Oxford University Press, 1962. 196
11. Maruhn, D. Rapid colorimetric assay of beta-galactosidase and N-acetyl-beta-glucosaminidase in human urine. *Clin Chim Acta* 1976 Dec; 73 (3) : 453-461
12. Jacobs W.L.W. a colorimetric assay for -glutamyl transpeptidase *Clin Chim Acta* 1971 Jan-Feb; 31 (1) : 175-179
13. Jung K. and Scholz D. An optimized assay of alanine aminopeptidase. *Clin Chem* 1980 Aug; 26 (9) : 1251-1254
14. Lee CY, Lee SY. Cardiovascular effects of snake venom. In : Lee CY, ed. *Snake Venoms.* Berlin : Springer-Verlay, 1979. 547
15. Vick JA, Ciucuta HP, Manthei JH. Pathophysiological studies of ten snake venoms. In : Russell FE, Saunder PR, eds. *Animal Toxins.* Oxford : Pergamon; 1967. 267
16. Huang HC. Effects of phospholipase A<sub>2</sub> from *Vipera russell* snake venom on blood pressure, plasma prostacyclin level and renin activity in rats. *Toxicon* 1984; 22 (2) : 254-264
17. Chaiyabutr N, Sitprija V, Kato S, Sugino N. Effect of converting enzyme inhibitor on renal function of rats following Russell's viper venom administration. *Jpn J Nephrol* 1985; (in press)
18. Zipser RD, Smorlesi C. Regulation of urinary thromboxane B<sub>2</sub> in man : Influence of urinary flow rate and tubular transport. *Prostaglandins* 1984 Feb; 27 (2) : 257-271
19. Chaiyabutr N, Sitprija V, Sugino N, Hoshi T. Russell's Viper Venom-Induced Depolarization in the Proximal Tubule of Triturus Kidney. *Thai J Vet Med* 1984 Dec; 15 (4) : 297-303
20. Thureau K, Boylan JW. Acute renal success. the unexpected logic of oliguria in acute renal failure. *Am J Med* 1976 Sep ; 61 (3): 308-315
21. Satoh H, Hosono M, Satoh S. Distinctive effect of angiotensin II on prostaglandin production in dog renal and femoral arteries. *Prostaglandins* 1984 Jun; 27 (6) : 807-820
22. McGiff JC, Crowshaw K, Terragno NA, Lonigro AJ. Release of prostaglandinlike substance into renal venous blood in response to angiotensin II. *Circ Res* 1970 Jul; 27 Supple 1 : 121
23. Blumberg AL, Denny SE, Marshall GR, Needleman P. Blood vessel-hormone interaction: angiotensin, bradykinin and prostaglandins. *Am J Physiol* 1977 Mar; 232 (3) : H305-H310
24. Piper PJ, Vane JR. Release of prostaglandins from lung and other tissues. *Ann N Y Acad Sci* 1971 ; 180 : 363

25. Zadik Z, Kowarski AA. Normal integrated concentration of aldosterone and plasma renin activity : effect of age. *J Clin Endocrinol Metab* 1980 May; 50 (5) : 867-869
26. Kunin Cm, Chesney RW, Craig WA, England AC, DeAngelis C. Enzymuria as a marker of renal injury and disease : studies of N-Acetyl- $\beta$  -glucosaminidase in the general population and in patients with renal disease. *Pediatrics* 1978 Nov; 62 (5) : 751-760
27. Gibey R, Dupond JL, Alber D, Leconte des Floris R, Henry JC. Predictive value of urinary N-acetyl-beta-D-glucosaminidase (NAG), alanine-aminopeptidase (AAP) and beta-2-microglobulin (beta<sub>2</sub>m) in evaluating nephrotoxicity of gentamicin. *Clin Chim Acta* 1981 Oct; 116 (1) : 25-34