

Procalcitonin and Interleukin-6 in pediatric patients admitted with suspected sepsis at **Tertiary Care Hospital**

Panipak Temboonnark* Kittipol Klaiklueng* Wissaroot Karoonboonyanan* Panjachat Ratanamongkol*

Temboonnark P, Klaiklueng K, Karoonboonyanan W, Ratanamongkol P. Procalcitonin and Interleukin-6 in pediatric patients admitted with suspected sepsis at Tertiary Care Hospital. Chula Med J 2018 Jan – Feb;62(1): 9 - 25

Background : Bacterial sepsis remains one of the leading causes of mortality and morbidity among children. Early identification of individuals at risk of developing life-threatening sepsis could enable early treatment and improve outcomes. Procalcitonin (PCT), the peptide hormone precursor of calcitonin, and Interleukin-6 (IL-6), the pro-inflammatory cytokine, are both substances that present in higher concentrations after inflammation. The potential value of measuring PCT and IL-6 levels are recognized as biological markers in pediatric patients with systemic inflammatory response syndrome (SIRS) in both early diagnosis of sepsis and to differentiate sepsis from the other SIRS non-septic conditions, has higher accuracy than the routine laboratory tests pragmatically such as total leukocytes count and C-reactive protein (CRP).

Objectives

: To define the clinical and laboratory predictive factors for early diagnosis in pediatric bacterial sepsis and distinguishing from the other SIRS non-septic conditions, including the development of the risk scoring system for pediatric bacterial sepsis prediction.

^{*}Department of Pediatrics, Bhumibol Adulyadej Hospital, Royal Thai Air Force

Methods

: The diagnostic prediction research was designed from the prospective data from September 2015 to June 2016 at the Department of Pediatrics, Bhumibol Adulyadej Hospital, Bangkok, Thailand. Forty-nine patients from the age of 1 month to 15 years old who met the criteria of SIRS definition were recruited into the study. Multivariable logistic regression was performed to select the strongest predictors then transformed to develop the final bacterial sepsis score (FBAC score).

Results

: The best clinical and laboratory predictors included female gender, PCT, IL-6, CRP levels prior to treatment on admission date and CRP level at 24 hours after starting of the treatment. The developed FBAC score predicted bacterial sepsis correctly with an AuROC of 86.5%. The patients who have FBAC scores below 10 (low risk category) will have the chance to be bacterial sepsis 10 times less (likelihood ratio of positive; LHR + 0.1, P - value 0.01) and the potential bacterial septic patients will have more than 6 times increasingly (LHR+ 6.13, P - value 0.02) if their total FBAC scores are higher than 30 (high risk category).

Conclusions: The developed FBAC scores will help pediatricians to predict bacterial sepsis for early treatment intervention and can distinguish bacterial sepsis from the other SIRS non-septic conditions.

Keywords

: Procalcitonin, Interleukin-6, systemic inflammatory response syndrome, bacterial sepsis, final bacterial sepsis score.

Correspondence to: Klaiklueng K. Department of Pediatrics, Bhumibol Adulyadej Hospital, Royal Thai Air Force, Bangkok 10220, Thailand.

Received for publication. April 5, 2017.

พาณิภัค เต็มบุญนาค, กิตติพล คล้ายคลึง, วิศรุต การุณบุญญานั้นท์, ปัญจฉัตร รัตนมงคล. โปรแคลซิโตนิน และอินเตอร์ลิวคิน 6 ในผู้ป่วยเด็กที่สงสัยภาวะติดเชื้อแบคทีเรีย ในกระแสเลือด. จุฬาลงกรณ์เวชสาร 2561 ม.ค. – ก.พ.;62(1): 9 – 25

เหตุผลของการทำวิจัย

: ภาวะติดเชื้อแบคทีเรียในกระแสเลือดเป็นสาเหตุสำคัญของภาวะ ทุพพลภาพ และการเสียชีวิตในผู้ปวยเด็ก ซึ่งถ้าผู้ปวยได้รับการวินิจฉัย และการรักษาอย่างรวดเร็วและเหมาะสมจะทำให้เกิดผลการรักษาที่มี ประสิทธิภาพมากขึ้น ปัจจุบันการวินิจฉัยภาวะติดเชื้อแบคทีเรียใน กระแสเลือด อาศัยการติดตามอาการทางคลินิกของผู้ปวยที่เข้าได้และ การเพาะเชื้อจากเลือด ซึ่งต้องใช้เวลาหลายวัน ดังนั้นการทำนาย ภาวะติดเชื้อแบคทีเรียในกระแสเลือดตั้งแต่ระยะเริ่มต้นก่อนการรักษา จึงมีความสำคัญเป็นอย่างยิ่ง โปรแคลซิโตนิน (procalcitonin) และ อินเตอร์ลิวคิน 6 (Interleukin-6) เป็นสารที่ถูกหลั่งออกมาเมื่อร่างกาย มีอาการตอบสนองต[่]อการอักเสบทั่วร่างกาย (systemic inflammatory response syndrome, SIRS) จึงสามารถช่วยในการวินิจฉัยภาวะ ติดเชื้อในกระแสเลือด และแยกความแตกตางระหวางผู้ปวยเด็ก ที่มีอาการแสดงของภาวะ SIRS ที่เกิดจากการติดเชื้อแบคทีเรียใน กระแสเลือดออกจากภาวะ SIRS ที่เกิดจากสาเหตุอื่นได้อยางรวดเร็ว และแม่นยำกว่าค่าทางห้องปฏิบัติการพื้นฐานที่ใช้ในปัจจุบัน เช่น ชีรีแอคทีฟโปรตีน (C-reactive protein) เป็นต[้]น เพื่อศึกษาอาการ อาการแสดงทางคลินิกและผลทางห้องปฏิบัติการ

วัตถุประสงค์

ที่ช่วยในการวินิจฉัยภาวะติดเชื้อแบคทีเรียในกระแสเลือดของผู้ป่วย เด็ก รวมทั้งศึกษาปัจจัยที่ช่วยแยกผู้ป่วยเด็กที่มีอาการแสดงของภาวะ SIRS ที่เกิดจากการติดเชื้อแบคทีเรียในกระแสเลือดออกจากภาวะอื่น ๆ

วิธีการทำวิจัย

SIRS ที่เกิดจากการติดเชื้อแบคทีเรียในกระแสเลือดออกจากภาวะอื่น ๆ

: เป็นการศึกษาเชิงวินิจฉัยและคาดคะเน (diagnostic prediction research) โดยเก็บข้อมูลไปข้างหน้า ระหว่างวันที่ 1 กันยายน 2558 ถึง 30 มิถุนายน 2559 ที่ห่อผู้ป่วยกุมารเวชกรรม และหออภิบาลผู้ป่วย กุมารเวชกรรม โรงพยาบาลภูมิพลอดุลยเดช กรมแพทย์ทหารอากาศ ในผู้ป่วยเด็กจำนวน 50 คน ซึ่งอายุระหว่าง 1 เดือนถึง 15 ปี ที่มีอาการ อาการแสดงทางคลินิกและผลทางห้องปฏิบัติการเข้าได้กับภาวะ SIRS โดยได้ส่งตรวจระดับโปรแคลซิโตนิน ระดับอินเตอร์ลิวคิน 6 และระดับ ซีรีแอคทีฟโปรตีนก่อนทำการรักษา หลังเริ่มการรักษา 24 ชั่วโมงและ หลังเริ่มการรักษา 72 ชั่วโมงตามลำดับ แล้วนำข้อมูลทั้งหมดมา วิเคราะห์โดยวิธี multivariate logistic regression เพื่อหาปัจจัย ทำนายที่มีความน่าเชื่อถือมากที่สุด (best predictive factors) แล้วนำ มาพัฒนาต่อเป็น final bacterial sepsis score (FBAC score) เพื่อใช้ ทำนายโอกาสเป็นภาวะติดเชื้อแบคทีเรียในกระแสเลือด

ผลการศึกษา

ข้อมูลอาการ อาการแสดงทางคลินิกและผลทางห้องปฏิบัติการที่มี ความน่าเชื่อถือในการทำนายภาวะติดเชื้อแบคทีเรียในกระแสเลือด ของผู้บ่วยเด็กมากที่สุด คือ เพศหญิง ระดับโปรแคลซิโตนินก่อนทำ การรักษา ระดับอินเตอร์ลิวคิน 6 ก่อนทำการรักษา ระดับซีรีแอคทีฟโปรตีนหลังเริ่มการรักษา ใปรตีนก่อนทำการรักษา และระดับซีรีแอคทีฟโปรตีนหลังเริ่มการรักษา 24 ชั่วโมง (P - value < 0.1 และ AuROC > 0.7) FBAC score มีค่า Area Under an ROC Curve (AuROC) เท่ากับร้อยละ 86.5 ค่า positive predictive value เท่ากับ ร้อยละ 71.4 และค่า negative predictive value เท่ากับ ร้อยละ 79.1 โดยผู้ป่วยที่มี FBAC score น้อยกว่าหรือ เท่ากับ 10 คะแนน (low risk category) มีโอกาสเป็นภาวะติดเชื้อ แบคทีเรียในกระแสเลือดลดลง 10 เท่า (likelihood ratio of positive; LHR + 0.1, P - value 0.01) แต่ถ้ามี FBAC score มากกว่าหรือเท่ากับ 30 คะแนน (high risk category) มีโอกาสเป็นการติดเชื้อแบคทีเรียในกระแสเลือดเพิ่มขึ้น 6 เท่า (likelihood ratio of positive; LHR + 6.13, P - value 0.02)

สรุป

จากงานวิจัยพบว่า FBAC score สามารถเป็นเครื่องมือในการทำนาย ภาวะติดเชื้อแบคทีเรียในกระแสเลือดของผู้ป่วยเด็กได้ ซึ่งอาจช่วยให้ กุมารแพทย์สามารถวินิจฉัยได้รวดเร็ว และสามารถให้การรักษา ตั้งแต่ระยะแรกได้อย่างมีประสิทธิภาพมากขึ้น นอกจากนั้นสามารถ ใช้แยกความแตกต่างระหว่างผู้ป่วยเด็กที่มีอาการแสดงของภาวะ SIRS ที่เกิดจากการติดเชื้อแบคทีเรียในกระแสเลือด ออกจากภาวะ SIRS ที่เกิดจากสาเหตุอื่นได้อย่างรวดเร็วและแม่นยำมากยิ่งขึ้น

คำสำคัญ

โปรแคลซิโตนิน, อินเตอร์ลิวคิน 6, กลุ่มอาการตอบสนองต่อการอักเสบ ทั่วร่างกาย, การติดเชื้อแบคทีเรียในกระแสเลือด, final bacterial sepsis score. Bacterial sepsis is one of the most common causes of mortality and morbidity among children especially in which severe sepsis with organ failure and septic shock which particularly profound circulatory, cellular and metabolic abnormalities associated with greater risk of mortality than with sepsis alone. Prompt and proper empirical antibiotic treatment is important for septic management. Blood culture is known as the gold standard test for diagnosis sepsis. However, its result hardly shows within 24 hours. (1) Total blood leukocytes count and C-reactive protein (CRP) level are universally used to be routine investigations for diagnosing sepsis but those still have limitations in accuracy and sometimes mislead the results. (2)

Procalcitonin (PCT) is the peptide precursor of calcitonin, a hormone that is synthesized by the parafollicular C cells of the thyroid gland and involved in calcium homeostasis. Normal serum PCT level is very low (<0.1 ng/ml). (3) PCT levels are obviously higher both in patients with sepsis and those with systemic inflammatory response syndrome (SIRS) from infection than the normal population. (4,5) Significantly, many studies have shown PCT is a biological marker for early diagnosis and as an indicator of severity among children with sepsis. (6,7) PCT is proved to be superior to CRP for diagnosis sepsis. (8-11) After inflammation, PCT level rises at 4hour interval (6 hours in CRP), peaking around 8 hours (36 - 50 hours in CRP) and takes 48 hours to reach normal level after appropriate treatment (72 - 96 hours in CRP). Furthermore, PCT also has more sensitivity and specificity for diagnosis sepsis, 92.6% and 97.5%, respectively. (12)

Interleukin-6 (IL-6) is an interleukin which acts as both pro-inflammatory cytokines and anti-inflammatory myokines, is secreted by T-cells and macrophages to stimulate immune response, e.g. during infection and after trauma, especially burns or other tissue damage leading to inflammation. (13, 14) Normally, IL-6 level is detected less than 5 pg/ml in the normal population but the level can rise within an hour after getting inflammation. (15) IL-6 is an early predictor for mortality in newly admitted patients who come with clinically SIRS. (16) IL-6 levels in septic shock patients are obviously higher than in sepsis patients. (17 - 20) In comparison with PCT and IL-6 levels are extremely higher at zero and twelve hours in septic patients with septic shock. (21) Moreover, some studies have shown PCT level is higher specificity whereas IL-6 is more sensitive and valuable for distinguishing bacteremia among oncohematologic patients with febrile neutropenia. (22)

According to neonatal sepsis has the unique complexity ^(23, 24), therefore the neonatal group is excluded from this study. Our hypothesis is that PCT and IL-6 levels are useful for early diagnosis of bacterial sepsis among children after neonatal period.

Objectives

The aim of our study is to define the clinical and laboratory predictive factors for early diagnosis in pediatric bacterial sepsis and distinguish sepsis from other SIRS non-sepsis, including the development of a scoring system for the prediction of pediatric bacterial sepsis.

Patients and Methods

Study design and participants

The diagnostic prediction research was designed from prospective data at the Pediatrics Department of Bhumibol Adulyadej Hospital, Royal Thai Air Force, Bangkok, Thailand during a 10-month period from 1 September 2015 to 30 June 2016.

The study has been approved by our Medical Research Institute and the Ethics Committee. After obtaining the informed consent, the pediatric patients between the age of 1 month and 15 years old who met the criteria of systemic inflammatory response syndrome (SIRS) definition were recruited into the study.

Diagnosis of SIRS and sepsis

According to the Pediatric Sepsis Consensus Congress 2015, systemic inflammatory response syndrome (SIRS) is defined as a patient having at least two of the following four criteria: core temperature of > 38.5 °C or < 36 °C, tachycardia (defined as mean heart rate > 2 SD above normal with the absence of external stimulus, chronic drugs, or painful stimuli, or otherwise unexplained persistent elevation over 0.5 -4 hours) or for children < 1 year: bradycardia (defined as mean heart rate < 10th percentile for age in the absence of external vagal stimulus, beta-blockers, congenital heart diseases, or otherwise unexplained persistent depression over 30 minutes), mean respiratory rate > 2 SD above normal of age or mechanical ventilator for an acute process not related to underlying neuromuscular disease or the receipt of general anesthesia and elevated or depressed total leukocyte counts for age (not secondary to chemotherapy-induced leukopenia) or > 10% immature neutrophils. One of these must be abnormal temperature or leukocyte count. (25)

Sepsis is defined when 2 or more SIRS criteria are present with a culture-proven or clinically identified infection. (25, 26) In our study, we used clinical follow-up data at the final course of treatment and /or positive hemoculture as the reference standard.

Patients with small cell lung carcinoma, medullary carcinoma of the thyroid gland, major trauma, cardiogenic shock and severe burn were excluded from this study.

Data collection and laboratory measurement

The clinical data of all recruited patients was collected including gender, age, underlying diseases, duration of fever, body temperature, blood pressure and heart rate.

The first blood sample was taken on admission date were complete blood count (CBC), blood urea nitrogen (BUN), creatinine, electrolytes, liver function test, blood culture, CRP (CRP-1), PCT (PCT-1), and IL-6 (IL-6-1). Subsequent two samples of CRP, PCT and IL-6 were taken at 24 hours (CRP-2, PCT-2, IL-6-2) and 72 hours later (CRP-3, PCT-3, IL-6-3). Resulting times of CRP, PCT, IL-6 were around 1 hour.

Blood volume at least 1 - 3 ml was taken for culture of all recruited patients before antibiotic was given. Blood culture was collected into BacT/Alert PF Plus Aerobic Pediatric bottle, delivered to the laboratory room not exceeding 1 hour, then incubated in Virtuo BacT/Alert and delivered into 5.0% blood agar plate and Macconkey agar plate.

CRP levels were measured by Roche P800 laboratory kit test, procalcitonin levels were measured by used Roche E170 laboratory kit test and IL-6 levels

were measured by Roche E601.

A value of PCT 0.5 to 2 ng/ml meant possible infection, 2 to 10 ng/ml indicated most likely sepsis and a value above 10 ng/ml showed severe bacterial sepsis or septic shock while IL-6 a value of IL-6 > 131 pg/ml referred to bacterial sepsis. (27,28) A plasma CRP of 50 mg/L or more was highly suggestive of sepsis. (29)

Statistical analysis

Clinical and laboratory data, as the potential predictors of sepsis obtained at time of admission included age, gender, underlying disease, onset of fever, body temperature, heart rate, shock status, total leukocytes count, absolute neutrophils count, band form neutrophils count, serum creatinine, serum total bilirubin, serum albumin, serum alanine aminotransferase (ALT), serum bicarbonate, serum CRP, serum PCT, serum IL-6, and hemoculture. Additionally, serum CRP, serum PCT and serum IL-6 were repeated at 24 and 72 hours later after starting treatment.

Two groups of patients, finally diagnosed from clinical follow-up data as sepsis and non-sepsis groups, were compared for evidence of differences (P - value) in all clinical and laboratory data with t - test, rank sum tests, chi-square or exact probability test as appropriate. Prediction by each clinical and laboratory data was calculated by univariate logistic regression analysis and presented as an area under the receiver operating characteristic (AuROC) curve. Strong (AuROC > 0.7) and significant (P - value < 0.1) clinical and laboratory data were categorized to facilitate odd ratio calculation, under multivariable logistic regression. Discriminative performance of the model was calculated by an AuROC curve. Regression

coefficients of each level of each predictor were divided by the smallest coefficient of the model to transform into an item risk score. Scores for each predictors were added up to obtain a total risk score, name final bacterial sepsis score (FBAC score). Score prediction of sepsis was done by using this FBAC score as the only summary predictor in the logistic model. Discrimination of the scores was presented with an AuROC curve. Calibration of the prediction was analyzed with Hosmer-Lemeshow statistics. Score predicting risks and observed risk were compared and presented in a graph as a risk curve. Risk scores were categorized into three risk levels, low, moderate and high. Predictive ability of each risk score level was calculated and presented as a likelihood ratio of positive (LHR+), 95.0% confidence interval and its significant level (P - value). Model and score performance are also expressed as sensitivity, specificity, predictive values and overall accuracy. All P - values < 0.05 were regarded as significant. Statistical analyses were done using STATA version 14.

It was estimated that a total sample of at least 40 patients would be adequate in our study. We chose the maximum sample size, calculated by two independent mean difference formula, by comparing all predictors among sepsis and non-sepsis SIRS patients, with a two tailed alpha level of 0.05 and a power of 90.0%, according to the 2 studies. (30, 31)

Results

A total of fifty patients, diagnosed with SIRS were included in the study. Twenty patients (40.0%) had final diagnosis with bacterial sepsis. The

univariate logistic analysis of all clinical characteristics and laboratory data are shown in Table 1 and 2. There were significantly female patients in sepsis group more than non–sepsis group (65.0% *vs* 35.0%, *P* - value 0.003). Other clinical characteristics as the age, underlying diseases of patients, duration of fever, shock status, body temperature, heart rate were not statistically different among sepsis and non-sepsis group. The laboratory data as total leukocytes count, absolute neutrophils count, band form neutrophils count, serum creatinine, serum bicarbonate, serum albumin, and serum alanine aminotransferase (ALT) were also not statistically different between the two groups.

Mean levels of all biomarkers on admission date prior to treatment, 24-hour interval and 72-hour interval after treatment between sepsis and non-sepsis

groups are shown in Table 3. In comparison, there were significant differences of PCT-1, PCT-2, PCT-3, IL-6-1, IL-6-2, IL-6-3, CRP-1 and CRP-2 levels between sepsis and non-sepsis groups (P - value < 0.1) and AuROC > 0.7. Among all clinical and laboratory predictors, the prediction ability as measured by the AuROC curve was highest for IL-6-1.

After the univariate logistic analysis, female gender, PCT-1, PCT-2, PCT-3, IL-6-1, IL-6-2, IL-6-3, CRP-1 and CRP-2 were chosen as the potential predictive factors due to P - value < 0.1 and AuROC > 0.7. The values of PCT-1, PCT-2, PCT-3, IL-6-1, IL-6-2, IL-6-3, CRP-1 and CRP-2 levels were categorized into 2 groups according to reference cut-off levels as shown in Table 4 to facilitate odd ratio calculation. The best predictors were then selected to performed multivariable logistic regression analysis.

Table 1. Clinical characteristics of sepsis cases vs. non sepsis cases, evidence of difference. (*P* - value) and area under the curve (AuROC)

Characteristics	Sepsis	Non sepsis	P	AuROC
	(n = 20)	(n = 30)		
Age (months); mean (95%CI)	59.4 (32.6 - 86.2)	57.3 (33.9 - 80.8)	0.91	0.51
Gender (female/male); n (%)	13 (65.0%) / 7(35.0%)	6 (20.0%) / 24 (80.0%)	0.01	0.73
Underlying disease; n (%)				
- Yes	7 (35.0%)	5 (16.7%)	0.37	0.60
- No	13 (65.0%)	25 (83.3%)		
Duration of fever(h); mean (95%CI)	64.6 (38.1 - 91.0)	71.7 (44.1 - 99.2)	0.71	0.50
Shock; n (%)	9 (45.0%)	5 (16.7%)	0.06	0.64
Body temp (°c); mean (95%CI)	38.5 (38.0 - 39.0)	38.4 (38.1 - 38.8)	0.86	0.55
Heart rate; mean (95%CI)	145 (134 - 158)	136 (127 - 146)	0.21	0.56

Table 2. Laboratory data on admission date of sepsis cases vs non sepsis cases, evidence of difference (*P* - value) and area under the curve (AuROC).

Laboratory data	Sepsis (n = 20)	Non-sepsis (n = 30)	P	AuROC
	Mean (95%CI)	Mean (95%CI)		
WBC count (cell/cumm³)	12,760.5 (6,361.6 - 19,159.4)	16,400.7 (12,565.7 - 20,235.6)	0.29	0.35
ANC (cell/cumm ³)	11,538.5 (4,296.1 - 18,780.9)	11,514.2 (7,915.8 - 15,112.5)	0.99	0.41
Band form neutrophils (%)	6.5 (3.0 - 10)	7.5 (4.4 - 10.6)	0.70	0.48
Creatinine (mg/dL)	0.4 (0.3 - 0.5)	0.4 (0.3 - 0.5)	0.92	0.52
Total bilirubin (mg/dL)	0.8 (0.3 - 1.3)	0.4 (0.3 - 0.4)	0.05	0.60
Serum albumin (g/dL)	3.1 (2.8 - 3.4)	4.6 (3.3 - 5.9)	0.06	0.12
AST (U/L)	58.0 (28.4 - 87.6)	34.0 (25.6 - 42.5)	0.07	0.61
ALT (U/L)	39.2 (20.4 - 57.9)	40.3 (23.6 - 57.0)	0.92	0.51
Serum bicarbonate (mEq/L)	17.2 (12.6 - 18.8)	23.2 (15.2 - 31.1)	0.21	0.45

WBC White blood cell, ANC Absolute neutrophil count, AST Aspartate aminotransferase, ALT Alanine- aminotransferase

Table 3. Laboratory data on admission date, 24 hours and 72 hours after admission of case sepsis and non sepsis, evidence of difference (*P* - value) and area under the curve (AuROC)

Laboratory data	Sepsis (n = 20) Non-sepsis (n = 30)		P	AuROC	
	Mean (95%CI)	Mean (95%CI)			
PCT-1 (ng/mL)	21.1 (7.8 - 34.5)	3.4 (1.6 - 5.2)	0.01	0.75	
PCT-2 (ng/mL)	20.4 (4.0 - 36.71)	6.0 (1.5 - 13.5)	0.07	0.77	
PCT-3 (ng/mL)	13.4 (-3.2 - 29.9)	1.4 (0.1 - 2.7)	0.07	0.70	
IL-6-1 (pg/mL)	682.8 (82.5 - 1,283.1)	49.7 (26.2 - 73.1)	0.01	0.86	
IL-6-2 (pg/mL)	193.0 (0.3 - 1.3)	34.1 (6.9 - 379.0)	0.03	0.83	
IL-6-3 (pg/mL)	86.3 (1.1 - 171.5)	22.6 (0.8 - 46.0)	0.08	0.77	
CRP-1 (mg/dL)	127.5 (73.8 - 181.3)	58.8 (28.6 - 89.0)	0.01	0.71	
CRP-2 (mg/dL)	127.6 (64.4 - 178.8)	43.3 (25.8 - 60.9)	0.01	0.80	
CRP-3 (mg/dL)	43.7 (16.4 - 70.9)	29.6 (10.2 - 49.0)	0.37	0.87	

PCT-1; PCT on admission date, PCT-2; PCT at 24 hours later, PCT-3; PCT at 72 hours later, IL-6-1; IL-6 on admission date, IL-6-2; IL-6 at 24 hours later, IL-6-3; IL-6 at 72 hours later, CRP-1; CRP on admission date, CRP-2; CRP at 24 hours later, CRP-3; CRP at 72 hours later

Table 4. Categorized laboratory data into groups according to the reference cut-off levels. (29)

Laboratory data	Sepsis	Non sepsis	P	AuROC
PCT-1 (ng/ml)				
< 2	6 (30.0%)	20 (66.7%)	0.02	0.68
≥2	14 (70.0%)	10 (33.3%)		
PCT-2 (ng/ml)				
< 2	6 (37.5%)	16 (66.7%)	0.11	0.65
≥2	10 (62.5%)	8 (33.3%)		
PCT-3 (ng/ml)				
< 2	8 (57.1%)	16 (80.0%)	0.25	0.61
≥2	6 (42.9%)	4 (20.0%)		
IL-6-1 (pg/ml)				
< 131	10 (50.0%)	26 (89.7%)	0.01	0.70
≥ 131	10 (50.0%)	3 (10.3%)		
IL-6-2 (pg/ml)				
< 131	10 (66.7%)	21 (87.5%)	0.22	0.60
≥131	5 (33.3%)	3 (12.5%)		
IL-6-3 (pg/ml)				
< 131	12 (85.7%)	19 (95.0%)	0.56	0.56
≥ 131	2 (14.3%)	1 (95.0%)		
CRP-1 (mg/dl)				
< 50	6 (31.6%)	19 (63.3%)	0.04	0.66
≥ 50	13 (68.4%)	11 (36.7%)		
CRP-2 (mg/dl)				
< 50	2 (13.3%)	15 (62.5%)	0.01	0.75
≥ 50	13 (86.7%)	9 (37.5%)		

After data categorization, the best 5 potential predictors remained female gender, PCT-1, IL-6-1, CRP-1 and CRP-2. They were chosen to make an exploratory model. An items score was assigned to each level of the 5 clinical and laboratory data by simple transformation of its logistic regression coefficient as shown in Table 5. The final bacterial sepsis score (FBAC score) was obtained by adding up the item scores.

The discriminative ability of the FBAC score which ranges from 0 to 37, could be observed by the

different percentage distribution between sepsis and non-sepsis groups as shown in Figure 1. The lower FBAC scores were more likely to be non-sepsis. On the other hand, the higher FBAC score were more likely to be sepsis. The FBAC score predicted bacterial sepsis with an AuROC of 86.5% as shown in Figure 2 and with the Hosmer-Lemeshow goodness of fit test of 0.55. In the risk curve showed that the scores predicted the risk of sepsis move upward when FBAC score increase, with close calibration to the actual or observed risks as shown in Figure 3.

Table 5. Best multivariable clinical predictors, odds ratio (OR), 95% confidence interval (CI), evidence of difference (*P* - value) and assigned item scores.

Predictors	OR	95% CI	P	Score
Gender				
Male	1.00	reference	-	0
Female	13.32	1.61 - 109.8	0.01	9.5
PCT on admission date (ng/ml)				
< 2	1.00	reference	-	0
≥ 2	1.31	0.05 - 10.0	0.83	1
IL-6 on admission date (pg/ml)				
< 131	1.00	reference	-	0
≥ 131	5.51	0.43 - 70.3	0.19	6
CRP on admission date (mg/dl)				
< 50	1.00	reference	-	0
≥ 50	6.46	0.01 - 3.0	0.22	7
CRP at 48 hours later (mg/dl)				
< 50	1.00	reference	-	0
≥ 50	38.78	1.35 - 1,107.0	0.03	13.5

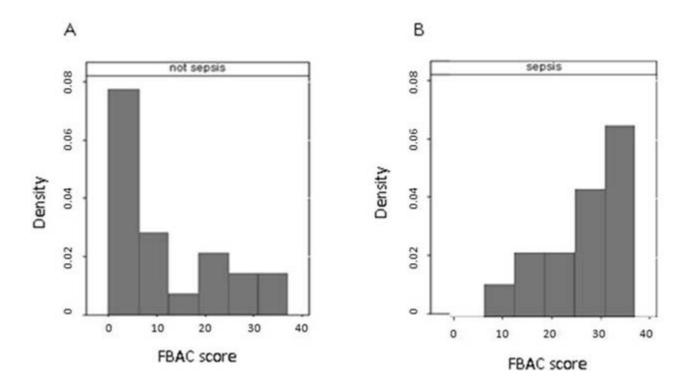


Figure 1. Percentage distribution of final bacterial sepsis (FBAC) score of A (not sepsis) and B (sepsis).

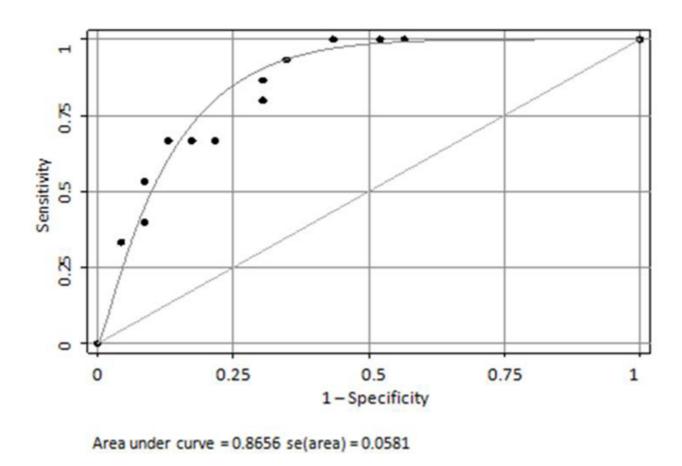


Figure 2. Area under receiver operating characteristic curve of FBAC scores on prediction of bacterial sepsis.

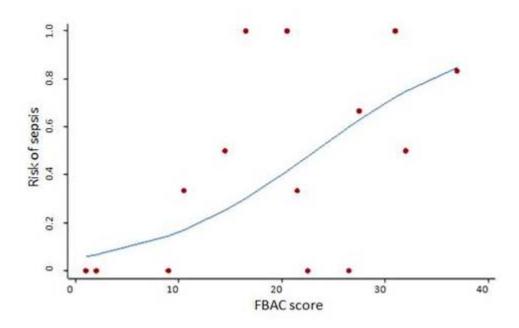


Figure 3. Observed risk (dot) vs. scores predicted risk (solid line) of bacterial sepsis.

The FBAC score were categorized into 3 risk groups, low (below 10) when the slope of the risk curve was lowest, moderate (10 to 30), and high (above 30) when the slope was highest to facilitate interpretation as shown in Table 5. At low category of FBAC score (below 10), the SIRS patients were significantly lower probability to be sepsis (LHR+ 0.10 and P - value 0.01) and at high category of FBAC score (above 30), the SIRS patients were significantly higher probability to be sepsis (LHR+ 6.13 and P - value 0.02).

The accuracy of FBAC score in distinguishing SIRS non-septic patients from septic patients is shown in Table 7. The score yielded a sensitivity of 66.67%, positive predictive value of 71.43%, of importance

the value reached to 82.61% and 79.17% in specificity and negative predictive value, respectively.

In this study, Only one patient had positive hemoculture. He was a boy, 7 months of age. His underlying disease were cerebral palsy and epilepsy. He was diagnosed as pneumonia with septic shock. His intial PCT, IL-6 and CRP were 5.8, 319.5 and 66.9 respectively. He received Piperacillin/Tazobactam. At 24 hour, his PCT, IL-6 and CRP became 2.8, 156.7 and 82.7 respectively. He died before 72 hours. His sputum culture was Stenotrophomonas maltophilia (MDR). The hemoculture, showed Pseudomonas aeruginosa (Extremly drug resisitant). The results of cultures returned after 3 days.

Table 6. Distribution of sepsis vs. non sepsis into low, moderate and high probability categories, likelihood ratio of positive (LHR+) and 95% confidence interval (CI).

Probability	FBAC	Seps	sis	Non	sepsis	LHR+	95% CI	<i>P</i> -value
categories	score	(n =	15)*	(n = 2	23)*			
		n	%	n	%			
Low	< 10	1	(6.7)	15	(65.2)	0.10	0.1 - 0.8	0.01
Moderate	10 - 30	6	(40.0)	6	(26.1)	1.53	0.3 - 6.9	0.52
High	> 30	8	(53.3)	2	(8.7)	6.13	1.0 - 64.6	0.02

LHR+: likelihood ratio of positive

Table 7. Sensitivity and specificity of final bacterial sepsis (FBAC) score.

	Final hasterial compic (FDAC) comp
	Final bacterial sepsis (FBAC) score
Sensitivity (%)	66.67%
Specificity (%)	82.61%
Positive predictive value (%)	71.43%
Negative predictive value (%)	79.17%
Accuracy (%)	76.31%

^{*}number of cases: with complete data

Discussion

The 50 pediatric patients who have been diagnosed with SIRS over 10-months period, there were 31 male (62.0%) and 19 female patients (38.0%) and male to female ratio is 1.63: 1 that are similar to other studies in other countries.⁽³²⁾

In the present studies show during the infectious process and sepsis, the effects of testosterone and androgen hormones can suppress the cell-mediated immune (CMI) response (immune depression). On the other hand, female hormones such as estrogen can promote the CMI response (immune modulation) that make the male is more likely to infect more often than female. (32) In our study, comparison between sepsis and SIRS non-sepsis groups, there are 13 female patients from 20 patients (65.0%) in sepsis group while there are 24 male patients from 30 patients in SIRS non-sepsis group (80.0%) this was distinguished from the past studies. Because this study was done with patients under the age of 15 years that is not under the reproductive age group, which has low level of estrogen; thus, there is no significant difference between males and females and no effect on female hormone levels to stimulate the CMI response.

Changes of leukocyte count, a routine laboratory test for sepsis, are common among septic patients. In fact, white blood cells > 12,000/cu.mm. or < 4,000/cu.mm. or shift to the left > 10.0% of immature neutrophils (bands) are one of the four SIRS criteria employed to diagnosis of sepsis but normal white blood cell count does not exclude sepsis, because many patients with sepsis have no leukocytosis, and many patients with leukocytosis have no infection. The white blood cell (WBC) count is

a not suitable test to determine infection and should be used only as part of validated multivariable decision rules for medical decision making. (33)

Prediction of bacterial sepsis has been a challenge of practice in pediatrics. It remains one of the leading causes of mortality and morbidity among children. Early identification of individuals at risk of developing life-threatening sepsis could enable early treatment and improve outcomes. In our study, the strongest potential clinical and laboratory predictors of sepsis are female gender, PCT concentration prior to treatment more than 2 ng/ml, IL-6 prior to the treatment more than 131 pg/ml, CRP prior to treatment and CRP at 24 hours after treatment more than 50 mg/dl. All these predictors can be assessed and the results obtained within one hour, therefore they can help pediatricians to predict probability of bacterial sepsis at the early course of treatment without waiting for blood culture or more blood sampling to determine.(34, 35)

Moreover, we have also developed simple final bacterial sepsis (FBAC) score. In our setting, the patients who scored below 10, will have a condition sepsis decreased 10 time (LHR + 0.1) and potential sepsis patients were more than six times the rate of total FBAC score more than 30 (LHR+ 6.13). The FBAC scores will be very useful for pediatrician to predict bacterial sepsis and differentiate between septic and SIRS non-septic conditions to make decision on early antibiotic treatment by using this score as the guideline.

Conclusion

The predictors of bacterial sepsis in SIRS patients from our study are female gender, high levels

of biomarker as PCT, IL-6 and CRP, the result of which could be obtained early in the course of treatment. A developed simple FBAC scores from all of the predictors will enable pediatricians to predict bacterial sepsis for early treatment intervention and can distinguish bacterial sepsis from the other SIRS nonseptic conditions.

Further studies should be conducted in larger numbers of pediatric patients and performed in multicenters to provide generalizability of the predictive model.

Acknowledgements

This study was supported by Bhumibol Adulyadej Hospital Fund. The authors have no conflict of interest to report. The authors, thereby, would like to offer our heartfelt thanks to Group Captain Nop Tuchinda, the chief of the Department Pediatrics, Bhumibol Adulyadej Hospital for the permission and valuable suggestions throughout the study.

This study was performed as part of the requirement for the diplomate, Thai Board of Pediatrics Medical Council of Thailand.

References

- Nanda SK, Dinakaran A, Bhat S, Ravichandran K, Kanungo R. Diagnostic and prognostic role of Procalcitonin in sepsis in a tertiary care hospital. Biomed Res 2016;27:79-83.
- 2. Mat-Nor MB, Md RA, Abdulah NZ, Pickering JW. The diagnostic ability of procalcitonin and interleukin-6 to differentiate infectious from noninfectious systemic inflammatory response syndrome and to predict mortality. J Crit Care 2016;33:245-51.

- 3. Fioretto JR, Borin FC, Bonatto RC, Ricchetti SM, Kurokawa CS, de Moraes M, et al. Procalcitonin in children with sepsis and septic shock. J Pediatr (Rio J) 2007;83: 323-8.
- 4. Biomarkers Definitions Working Group. Biomarkers and surrogate endpoints: preferred definitions and conceptual framework. Clin Pharmacol Ther 2001;69:89-95.
- 5. Schelonka RL, Infante AJ. Neonatal immunology. Semin Perinatol 1998;22:2-14.
- 6. Carrol ED, Thomson AP, Hart CA. Procalcitonin as a marker of sepsis. Int J Antimicrob Agents 2002;20:1-9.
- 7. Casado-Flores J, Blanco-Quiros A, Asensio J, Arranz E, Garrote JA, Nieto M. Serum procalcitonin in children with suspected sepsis: a comparison with C-reactive protein and neutrophil count. Pediatr Crit Care Med 2003:4:190-5.
- 8. Sherwin C, Broadbent R, Young S, Worth J, McCaffrey F, Medlicott NJ, et al. Utility of interleukin-12 and interleukin-10 in comparison with other cytokines and acute-phase reactants in the diagnosis of neonatal sepsis.

 Am J Perinatol 2008;25:629-36.
- 9. Simon L, Saint-Louis P, Amre DK, Lacroix J, Gauvin F. Procalcitonin and C-reactive protein as markers of bacterial infection in critically ill children at onset of systemic inflammatory response syndrome. Pediatr Crit Care Med 2008;9:407-13.
- 10. Sakha K, Husseini MB, Seyyedsadri N. The role of the procalcitonin in diagnosis of neonatal sepsis and correlation between procalcitonin

- and C-reactive protein in these patients.

 Pak J Biol Sci 2008;11:1785-90.
- 11. Meisner M, Adina H, Schmidt J. Correlation of procalcitonin and C-reactive protein to inflammation, complications, and outcome during the intensive care unit course of multiple-trauma patients. Crit Care 2006;10: R1.
- 12. Andreola B, Bressan S, Callegaro S, Liverani A, Plebani M, Da Dalt L. Procalcitonin and Creactive protein as diagnostic markers of severe bacterial infections in febrile infants and children in the emergency department. Pediatr Infect Dis J 2007;26:672-7.
- 13. Enguix A, Rey C, Concha A, Medina A, Coto D, Dieguez MA. Comparison of procalcitonin with C-reactive protein and serum amyloid for the early diagnosis of bacterial sepsis in critically ill neonates and children. Intensive Care Med 2001;27:211-5.
- 14. Magudumana MO, Ballot DE, Cooper PA, Trusler J, Cory BJ, Viljoen E, et al. Serial interleukin 6 measurements in the early diagnosis of neonatal sepsis. J Trop Pediatr 2000;46:267-71.
- 15. Reinhart K, Meisner M, Brunkhorst FM. Markers for sepsis diagnosis: what is useful? Crit Care Clin 2006;22:503-x.
- 16. Goldstein B, Giroir B, Randolph A. International pediatric sepsis consensus conference: definitions for sepsis and organ dysfunction in pediatrics. Pediatr Crit Care Med 2005;6: 2-8.
- 17. Yulianto S, Runtunuwu AL, Pateda V, Mandei JM, Lolombulan JH. Correlation between

- interleukin-6 and septic shock in children. Paediatr Indones 2012;52:352.
- 18. Pavare J, Grope I, Kalnins I, Gardovska D. High-mobility group box-1 protein, lipopolysaccharide-binding protein, interleukin-6 and C-reactive protein in children with community acquired infections and bacteraemia: a prospective study. BMC Infect Dis 2010;10:28.
- 19. Fioretto JR, Martin JG, Kurokawa CS, Carpi MF, Bonatto RC, Ricchetti SM, et al. Interleukin-6 and procalcitonin in children with sepsis and septic shock. Cytokine 2008;43:160-4.
- 20. Briassoulis G, Venkataraman S, Thompson A. Cytokines and metabolic patterns in pediatric patients with critical illness. Clin Dev Immunol 2010;2010:354047.
- 21. Luzzani A, Polati E, Dorizzi R, Rungatscher A, Pavan R, Merlini A. Comparison of procalcitonin and C-reactive protein as markers of sepsis. Crit Care Med 2003;31: 1737-41.
- 22. Urbonas V, Eidukaite A, Tamuliene I. The diagnostic value of interleukin-6 and interleukin-8 for early prediction of bacteremia and sepsis in children with febrile neutropenia and cancer. J Pediatr Hematol Oncol 2012; 34:122-7.
- 23. Wynn JL, Wong HR. Pathophysiology and treatment of septic shock in neonates. Clin Perinatol 2010;37:439-79.
- 24. Dellinger RP, Levy MM, Rhodes A, Annane D, Gerlach H, Opal SM, et al. Surviving Sepsis Campaign: international guidelines for management of severe sepsis and septic

- shock, 2012. Intensive Care Med 2013;39: 165-228.
- 25. Mathias B, Mira JC, Larson SD. Pediatric sepsis.

 Curr Opin Pediatr 2016;28:380-7.
- 26. Standage SW, Wong HR. Biomarkers for pediatric sepsis and septic shock. Expert Rev Anti Infect Ther 2011;9:71-9.
- 27. Meisner M. Procalcitonin-Biochemistry and Clinical Diagnosis. Bremen: Uni-Med Verlag Ag; 2010.
- 28. American College of Chest Physicians/Society of Critical Care Medicine Consensus Conference: definitions for sepsis and organ failure and guidelines for the use of innovative therapies in sepsis. Crit Care Med 1992;20:864-74.
- 29. Povoa P, Almeida E, Moreira P, Fernandes A, Mealha R, Aragao A, et al. C-reactive protein as an indicator of sepsis. Intensive Care Med 1998;24:1052-6.
- 30. Balci C, Sivaci R, Akbulut G, Karabekir HS.

 Procalcitonin levels as an early marker in patients with multiple trauma under intensive care. J Int Med Res 2009;37:1709-17.

- 31. Pavare J, Grope I, Eihvalde L, Gardovska D.

 Diagnostic markers for identifying sepsis
 in patients with Systemic Inflammatory
 Response Syndrome (SIRS): A prospective
 study. Open Pediatr Med J 2009;3:1-7.
- 32. Angele MK, Pratschke S, Hubbard WJ, Chaudry IH. Gender differences in sepsis: cardiovascular and immunological aspects. Virulence 2014;5:12-9.
- 33. Seigel TA, Cocchi MN, Salciccioli J, Shapiro NI, Howell M, Tang A, et al. Inadequacy of temperature and white blood cell count in predicting bacteremia in patients with suspected infection. J Emerg Med 2012;42: 254-9.
- 34. Samraj RS, Zingarelli B, Wong HR. Role of biomarkers in sepsis care. Shock 2013;40: 358-65.
- 35. Yang Y, Xie J, Guo F, Longhini F, Gao Z, Huang Y, et al. Combination of C-reactive protein, procalcitonin and sepsis-related organ failure score for the diagnosis of sepsis in critical patients. Ann Intensive Care 2016;6:51.